

Immune cell counts and risks of respiratory infections among infants exposed pre- and postnatally to organochlorine compounds: a prospective study

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Abstract

Background: Early-life chemical exposure may influence immune system development, subsequently affecting child health. We investigated immunomodulatory potentials of polychlorinated biphenyls (PCBs) and *p,p'*-DDE in infants.

Methods: Prenatal exposure to PCBs and *p,p'*-DDE was estimated from maternal serum concentrations during pregnancy. Postnatal exposure was calculated from concentrations of the compounds in mother's milk, total number of nursing days, and percentage of full nursing each week during the 3 month nursing period. Number and types of infections among infants were registered by the mothers (N=190). White blood cell counts (N=86) and lymphocyte subsets (N=52) were analyzed in a subgroup of infants at 3 months of age.

Results: Infants with the highest prenatal exposure to PCB congeners CB-28, CB-52 and CB-101 had an increased risk of respiratory infection during the study period. The odds ratio (OR) for infections decreased with increasing prenatal mono-*ortho* PCB exposure (CB-105, CB-118, CB-156, CB-167) and postnatal di-*ortho* PCB (CB-138, CB-153, CB-180) exposure. Similarly, infants with the lowest prenatal di-*ortho* PCB exposure and postnatal mono-*ortho* PCB and *p,p'*-DDE exposure, had a higher OR, than infants with higher exposures. Altogether, a negative relationship was indicated between infections and total organochlorine compound exposure during the whole pre- and postnatal period. Prenatal exposure to CB-28, CB-52 and CB-101 was positively associated with numbers of lymphocytes and monocytes in infants 3 months after delivery. A negative association between prenatal exposure to mono- and di-*ortho* PCBs and percentage of CD8⁺ cytotoxic T-cells was observed, whereas prenatal exposure to *p,p'*-DDE was negatively associated with the percentage of eosinophils.

Conclusions: This hypothesis generating study suggest that background exposure to PCBs and *p,p'*-DDE early in life modulate immune system development. Strong correlations between

mono- and di-*ortho* PCBs, and *p,p'*-DDE exposures make it difficult to identify the most important contributor to the suggested immunomodulation, and to separate effects due to pre- and postnatal exposure. Our findings may have consequences for the health development during childhood, since respiratory infections early in life may be risk factors for asthma and middle ear infections.

Background

Persistent and lipophilic organochlorine compounds, such as the industrial chemicals polychlorinated biphenyls (PCBs), the pesticide DDT, and dioxin-like contaminants polychlorinated dibenzo-*p*-dioxins (PCDDs) and dibenzofurans (PCDFs), are immunotoxic to animals and humans [1-7]. Suppression of the humoral and cellular immune system is one of the most sensitive endpoints after prenatal exposure of animals to the highly toxic dioxin-like compound 2,3,7,8-tetrachloro dibenzo-*p*-dioxin (TCDD) [6,7]. Children from Taiwan, who were accidentally exposed to high levels of both non-dioxin-like and dioxin like PCBs and polychlorinated dibenzofurans (PCDFs) prenatally, had higher rates of bronchitis, upper respiratory infections and middle ear infections than normally found in reference populations with low exposure [4,5].

A few studies have reported that exposure of the fetus and/or infant to background levels of non-dioxin-like and dioxin-like PCBs, the DDT metabolite *p,p'*-DDE, and PCDD/Fs are associated with alterations in markers of immune function, such as white blood cell (WBC) counts and numbers of lymphocyte subsets during childhood [8-13]. Diverging results have however been reported. On one hand no associations were found between WBC/lymphocyte subset counts and early life exposure to PCBs among Inuit infants [13]. On the other hand, among Dutch infants negative associations were found between monocyte counts and PCB exposure, and positive associations between CD8⁺ cytotoxic T-cells and PCB exposure [12].

High prenatal PCB exposure has been associated with a decreased thymus size among neonates born in an area with high environmental load of both non-dioxin-like and dioxin-like PCBs in Eastern Slovakia [14]. PCB exposure early in life has been positively associated with acute *otitis media* and negatively associated with incidence of asthma or allergies later in life in children [10,11,15]. Studies of preschool and school children have also suggested that exposure

to *p,p'*-DDE early in life may be a risk factor for acute otitis media and asthma later during childhood [13,16,17]. A decreased antibody response after tetanus and diphtheria vaccination was found among children exposed to high levels of PCBs early in life [18].

In order to better understand how PCB compounds and *p,p'*-DDE may influence children health, we studied the associations between pre- and postnatal PCB and *p,p'*-DDE exposure and numbers and percentages of WBCs and lymphocyte subsets in three month old infants. Moreover, associations between organochlorine compound exposure and upper/lower respiratory infections in infants were studied during the first three months of life. Compounds were assigned to one of four compound groups depending on biological activity and sources of exposure: non-dioxin-like tri- to pentachlorinated PCBs (CB-28, CB-52 and CB-101), non-dioxin-like di-*ortho* PCBs (CB-138, CB-153 and CB-180), dioxin-like mono-*ortho* PCBs (CB-105, CB-118, CB-156 and CB-167), and *p,p'*-DDE.

Methods

Study population and sampling. Between 1996 and 1999, 325 primiparous women, living and seeking prenatal care in Uppsala County, Sweden, agreed to donate a serum sample (82% participation rate) in late pregnancy (week 32-34) for chemical analysis of organochlorines [19]. PCB and DDE concentrations in serum lipids from this sampling were used as an estimate of prenatal PCB and DDE exposure of the infants, and 190 mother/child pairs had complete data for statistical analyses of associations between prenatal PCB and DDE exposure and diseases among the infants (Table 1).

During the third week after delivery mothers from the group of 190 participants sampled milk while nursing their infants, using a manual breast pump and/or a passive mother's milk

sampler [20]. The results of the PCB and DDE analyses of mother's milk was used in the assessment of postnatal exposure (see below) and 175 mother/infant pairs had complete data for statistical analyses of associations between postnatal PCB and DDE exposure and risk of infections.

Within this group of 175 mother/infant pairs, blood was sampled from 86 infants 3 months after delivery for analysis of numbers and percentages of WBCs and lymphocyte subsets. The age of 3 months was chosen in order to facilitate comparison with other studies of immunomodulatory effects of early life PCB and DDE exposure [11,13]. Moreover, the majority of the infants in Sweden have mothers' milk as the only source of nutrition during the first 3 months of life, making confounding of results due to differences in source of nutrition less pronounced. Blood samples could not be taken from the whole group of 175 infants due to lack of financial resources. All mothers were asked if they were willing to donate an infant blood sample, and all infants of mothers accepting were sampled until the number of 86 samples was reached. The participation rate in the blood sampling of the infants was 49%. Among these 86 mother/infants pairs data on numbers of WBCs were finally available from 81 infants and percentage data from 85 infants. Lymphocyte subsets could only be analyzed in 52 infants from this subgroup of 86 infants, due to limited volumes of blood available. Among the infants with data on lymphocyte subsets, data on lymphocyte numbers and percentages were available for 47 and 52 infants, respectively. The study was approved by the Ethics Committee of the Medical Faculty at Uppsala University. All participating women gave their informed consent prior to the inclusion in the study group.

Interviews and questionnaires. At 6-12 and 32-34 completed gestational weeks, in-person interviews regarding maternal characteristics were performed, using a structured questionnaire

[19,21]. Data on maternal characteristics included age, height, body weight before pregnancy, body weight at interview, years of education, and alcohol consumption and smoking before and during pregnancy. Blood samples for cotinine analysis (used as indicator of smoking habits) were taken at both interview occasions.

After delivery, the mothers were visited by a midwife when the infant was 3 months old. In an in-person interview, using a structured questionnaire, the mothers answered questions about sex of the infant, vaccination, nursing habits, and infant health during the first 3 months. Women gave information about the extent of nursing for each week of the 3 month period (full nursing, partial nursing and no nursing). The participants also gave information about the health of the children each week from delivery to the date of the interview. It was asked if the infant had any infection or other disease during the 3 month period. If the answer was yes, the mother was asked to give information about type of disease (open ended question). The mother was asked to identify the week/weeks the infant had the disease in an almanac, and to give the number of days the symptoms lasted. It was also asked if the infant had a fever during these days, and number of days with a temperature above 39⁰C. Finally it was asked if the symptoms were treated by a physician and what type of treatment that was used. Both the interviewer and the mothers were blinded to the organochlorine compound body burdens of the mother.

Organochlorine compound analysis. The lipid portion of serum samples was analyzed for the DDT metabolite *p,p'*-DDE, and 10 PCB congeners (IUPAC nos. 28, 52, 101, 105, 118, 138, 153, 156, 167 and 180). The analytical results were consequently lipid adjusted. Procedures for extraction, sample clean-up and analysis, and quality control are described in Glynn et al. [19]. In mother's milk the compounds were analyzed using a method and quality assurance described by

Glynn et al. [20]. When concentrations were below the limit of quantification (LOQ) they were set to 50% of LOQ in the statistical analysis.

Immune cell analysis. The hematological tests were performed at the Department of Clinical Chemistry, University Hospital of Uppsala. Capillary blood samples were collected in microtainer tubes (Sarstedt, Sweden). Differential counts were carried out by an automated instrument, Celldyn 4000 (Abbott Scandinavia AB, Solna, Sweden) based on a combination of optical characteristics and histochemical reactions. Number of total WBCs, and number and percentages of neutrophils, eosinophils, lymphocytes, and monocytes were recorded.

Analysis of lymphocyte subpopulations was performed on mononuclear cells prepared by Ficoll-Paque centrifugation and stained as described in Gräske et al. [22]. The following subpopulations were evaluated by flow cytometry: CD3⁺ cells (T-lymphocytes), CD19⁺ cells (B-lymphocytes), CD4⁺ cells (T-helper cells), CD8⁺ cells (cytotoxic cells), and CD56⁺ cells (NK-cells).

Calculations and statistics. Lipid adjusted serum organochlorine compound concentrations of the mothers in late pregnancy were used as a measure of prenatal exposure of the fetus [23-25].

Postnatal exposure (PE) (ng or pg/g*days) was calculated from the organochlorine concentration (ng or pg/g fresh weight) in mother's milk three weeks after delivery (Oconc), the number of days of nursing (Nd), and the percentage of full nursing during the study period (%N) using the equation

$$PE \text{ (ng or pg/g*days)} = Oconc * Nd * (\%N/100).$$

For women who did not nurse their infants PE=0. For the calculation of post-natal organochlorine exposure, organochlorine compound concentrations on a fresh weight basis were used since this more correctly represents the amount of the compounds that is ingested by the infant during nursing.

In the exposure analysis, tri- to penta-chlorinated CB-28, CB-52 and CB-101, with no dioxin-like biological activity, were grouped together (CB 28+52+101) because they showed low correlations with levels of the other organochlorine compounds [19]. These three congeners have been detected in elevated levels in indoor air and in the blood of residents of buildings containing PCB-laden building materials [26-29]. CB-138, CB-153 and CB-180 (di-*ortho* PCBs) were grouped together because of high correlations between serum concentrations of these congeners [19], and because of their lack of dioxin-like biological activity [30]. Dioxin-like mono-*ortho* PCBs consisted of CB-105, CB-118, CB-156 and CB-167 [30]. *p,p'*-DDE was treated separately in the statistical analysis.

Statistical analysis was performed using MINITAB® For Windows, 14. Spearman's rank correlation analysis was used in analysis of correlations between different exposure variables. The association between immune cell numbers/percentages and organochlorine compound exposure was explored by linear regression analysis. Regression analysis was performed on logarithmically transformed organochlorine compound exposure data, since the distributions of data closely followed a log-normal distribution. However, in the case of prenatal exposure to CB 28+52+101 many women had serum levels below LOQ and this exposure variable was therefore categorized. Some women did not nurse their infants at all, and postnatal exposure of these infants was set to zero. This made logarithmic transformation impossible and postnatal exposure was therefore categorized. Prenatal CB 28+52+101 exposure was categorized in 3 categories. The study participants could not be categorized in tertiles since over 40% had concentrations below

the LOQ. Therefore mother/infants pairs with CB 28+52+101 concentrations below the LOQ in were grouped the reference category and the rest of the study participants divided up in equal numbers in the two other categories depending on exposure levels. Postnatal organochlorine compound exposure was categorized in quartiles when possible, otherwise an effort was made to have equal numbers of study participants in each of the four exposure categories.

In the statistical analysis of immune cell results the significance level was set to $p \leq 0.01$, since multiple comparisons were made. In cases when a statistically significant association between immune cells and organochlorine compound exposure was found in simple regression analysis, multiple regression analysis was used to adjust the associations for potential confounders. We included lifestyle/medical factors in the multiple regression analyses that have previously been reported to be associated with increased or decreased risk of immune-related diseases [31-36]. Potential confounders included in the analysis were age of the mother, smoking during pregnancy (non-smoker/former smoker/smoked during pregnancy), alcohol consumption during pregnancy (no/yes), mother's education (≤ 13 years of education/14-16 years of education/ > 16 years of education), vaccination status of the infant at the time of sampling (yes/no), nursing of the infant (full nursing/no and partial nursing), age of the infant, and infant respiratory infections during the period before sampling (no/yes). In linear regression analysis a few observations with a standardized residual ≥ 3.0 were omitted from the data sets due to a large impact on the regression results.

In the analysis of infection results (no/yes), odds ratios (ORs) and 95% confidence intervals (CIs) were calculated using logistic regression. Respiratory infection (influenza-like symptoms, common cold, or common cold with cough) (N=54) was the only health problem that was frequent enough to allow statistical analysis. Other health problems reported were stomach problems (N=1), navel infection (N=1), cows' milk intolerance/allergy (N=1), urinary infection

(N=1), eye infection (N=2) and middle ear infection (N=2). In the regression analysis all independent variables were categorized in order to handle outlier problems. First, ORs were calculated in regressions with the exposure to each organochlorine compound group as the only independent variable. In the next step the ORs were adjusted for the same possible confounders as in the analyses of immune cell counts/percentages (except infants' infection, see above). A stricter definition of respiratory infection was also used (three days or more of infection, N=50) in the statistical analysis. The information about number of infections during the study period could not be used in the statistical analyses of the results, since only 7 infants had more than one infection during the study period. Similarly, too few infants had been examined by a physician. It was not possible to use the information about infection in combination with information about body temperature in the statistical analysis, since there were too many missing values for the latter variable.

We also analyzed the associations between respiratory infections and the total exposure to PCBs and *p,p'*-DDE during the whole prenatal and postnatal period. This analysis was based on a summation of the categorized pre- and postnatal exposure to all compound groups. We did not calculate a sum of the absolute concentrations since such a summation assumes equal immunomodulatory potency of all the studied compounds. For example, when absolute concentrations are summed up, effects of relatively potent compounds present at low concentrations may be underestimated, and effects of less potent compounds present at high concentrations overestimated. We avoided this by summing up exposure scores for each compound group. First the pre- and postnatal PCB and *p,p'*-DDE exposures were separately scored, with the lowest score 1 given to the lowest exposure quartile and the highest score 4 given to the highest exposure quartile (scores 1-3 for prenatal CB 28+52+101 exposure). The scores for pre- and postnatal exposures were then summarized for each compound group. For

example, if an infant scored 1 for prenatal exposure to *p,p'*-DDE and scored 3 for postnatal *p,p'*-DDE exposure then the infant was assigned a score of 4 in the summation of pre- and postnatal *p,p'*-DDE exposure. The resulting sum of the pre- and postnatal exposure scores of each compound group was then added to get the total exposure score. In this case, if an infant had scored 5 for total CB 28+52+101 exposure, 8 for mono-*ortho* PCB exposure, 6 for di-*ortho* PCB exposure, and 5 for *p,p'*-DDE exposure, the total organochlorine compound exposure score was summed up to 24. The total exposure scores were subsequently divided into quartiles in the final statistical analyses of associations between infections and total organochlorine exposure. The level of statistical significance in logistic regression analysis was set to $p \leq 0.05$.

Results

Personal characteristics. Table 1 gives the personal characteristics of the whole study group (infection), the subgroup of mother/infant pairs that donated blood for WBC counts, and mother/infant pairs within this subgroup that donated enough blood for lymphocyte subset analyses. A comparison of personal characteristics between the study participants that did not donate infant blood (N=104) with those donating infant blood (N=86) showed no significant differences in age of the mothers and infants (t-test, $p \geq 0.05$) (Table 1). No difference was found between the two subgroups in smoking during pregnancy, alcohol consumption during pregnancy, years of education, nursing, and respiratory infections (Chi-square test, $p \geq 0.05$). Furthermore, pre- and postnatal organochlorine compound exposure did not differ between the groups (t-test, $p \geq 0.05$). However, there were more girls than boys in the subgroup with blood samples, whereas the reverse was evident among mother/infant pairs that did not donate infant

blood (Chi-square test, $p \leq 0.05$). Fewer had been vaccinated at the end of the study period among infants with blood samples (Chi-square test, $p \leq 0.05$).

Pre- and postnatal exposure. Among the PCB congeners, the median prenatal exposure of the sum of CB-28, CB-52 and CB-101 was low (Table 2). Over 40% of the mothers in the whole study group had concentrations of all the three substances in serum lipids below the LOQ. Concentrations ≥ 10 ng/g lipid were however found in 15% of the study participants. Median concentrations of di-*ortho* PCBs (CB-138, CB-153 and CB-180) and *p,p'*-DDE were similar, and the concentrations varied 8- to 26-fold. The concentrations of mono-*ortho* PCB TEQs varied 9-fold or more (Table 2).

Correlations between serum concentrations of CB 28+52+101 and the other organochlorines in the total study group (infection) were weak, with Spearman correlation coefficients ranging from 0.13 to 0.24. Concentrations of di-*ortho* PCBs and mono-*ortho* PCB TEQs were strongly correlated ($r=0.93$), whereas correlations between these two PCB groups and *p,p'*-DDE were less strong ($r=0.66-0.75$).

Variation in organochlorine exposure from mother's milk during the first 3 months of infancy was large (Table 2). The correlation between postnatal exposure to di-*ortho* PCBs and mono-*ortho* TEQs was strong ($r=0.95$). Other correlations between the studied compound groups were less strong ($r=0.40-0.82$). Pre- and postnatal exposure of each compound group was significantly correlated, ranging from $r=0.75$ to $r=0.78$.

Organochlorine compound exposure and WBCs. The numbers and percentages of WBCs are presented in Table 3. In both simple and multivariate regression analysis, infants with the highest CB 28+52+101 exposure had significantly higher mean numbers of total WBCs, lymphocytes

and monocytes than infants in the reference category with the lowest exposure (Figure 1). Even after exclusion of infants with ongoing infection at the time of sampling (N=6), infants with the highest CB 28+52+101 exposure had higher WBC counts (regression coefficient: 2.69 ± 0.49 , $p \leq 0.01$) than infants with the lowest exposure. The results for lymphocytes and monocytes did either not change when infants with an ongoing infection were excluded (lymphocyte: 1.78 ± 0.40 , $p \leq 0.01$; monocyte: 0.13 ± 0.05 , $p \leq 0.01$). The percentage of eosinophils was negatively associated with prenatal *p,p'*-DDE exposure (Table 4). Also in this case the results did not change after exclusion of infants with ongoing infection at time of sampling (regression coefficient: -1.62 ± 0.46 , $p \leq 0.01$). Otherwise, no significant associations between prenatal exposure and WBC counts/percentages were found (Figure 1, Table 4).

Numbers and percentages of WBCs were not significantly associated with postnatal exposure to any of the analyzed organochlorines (see additional files: Table A and Table B).

Organochlorine exposure and lymphocyte subsets. We found a statistically significant negative association between percentage of CD4⁺CD8⁺ cells and prenatal exposure to di-*ortho* and mono-*ortho* PCBs in the univariate and multivariate analyses (Table 4). The results did not change when infants with ongoing infection at time of sampling were excluded, (mono-*ortho* PCBs regression coefficient: -5.57 ± 2.06 , $p \leq 0.01$; di-*ortho* PCBs: -4.49 ± 2.06 , $p \leq 0.05$). Otherwise numbers and percentages of different lymphocyte subsets were not associated with prenatal exposure (Figure 2, Table 4). Postnatal exposure to the organochlorines was not significantly associated with numbers and percentages of lymphocyte subsets (see additional file: Table C and Table D).

Respiratory infections. Infants with the highest prenatal CB 28+52+101 exposure had a significantly increased odds ratio for respiratory infections during the first three months after birth compared with the lowest exposed infants (Table 5). This finding did not change in multivariate-adjusted logistic regression analysis. Moreover, a lowered odds ratio was found for infants in the second to fourth exposure category of prenatal di-*ortho* PCB exposure, although the difference between the highest exposure category and the reference category only showed borderline statistical significance (Table 5). Similarly, a lowered odds ratio was found for infants in the highest exposure category of prenatal mono-*ortho* PCB exposure in the multivariate-adjusted analysis (Table 5). Infants with the lowest prenatal *p,p'*-DDE exposure had a higher odds ratio than infants with higher *p,p'*-DDE exposures, but the difference never reached statistical significance. Similar results were evident when a stricter definition of respiratory infection (3 days or more of infection) was used (see additional file: Table E).

In the analysis of postnatal exposure, the OR for respiratory infections among infants in the highest exposure category of di-*ortho* PCB exposure was significantly lower than that of the reference category in the unadjusted analysis (Table 5). Multivariate analysis did not change this difference markedly. Significantly lowered ORs were observed for the second exposure category of *p,p'*-DDE and mono-*ortho* PCB, suggesting a U-shaped relationship (Table 5). An U-shaped relationship was also suggested for CB 28+52+101, but the differences in odds ratio between the reference category and the other exposure categories were not statistically significant. Similar results were found when infections lasting less than 3 days were excluded in the definition of infections (see additional file: Table E).

ORs for respiratory infections among infants in the second, third and fourth categories of total organochlorine compound exposure during the whole pre- and postnatal period were 0.74 (95% confidence interval:0.29-1.88), 0.36 (0.13-0.96), and 0.68 (0.28-1.68), respectively.

Discussion

In this exploratory study of associations between early life exposure to organochlorine compounds and risks of respiratory infections among 3 month old infants, diverging associations between exposure to organochlorines compounds and infection risk were found. Our results suggest that high prenatal background exposure to the PCB congeners CB-28, CB-52 and CB-101 may increase the risk of respiratory infections among infants. The reverse was suggested among infants with high prenatal exposure to mono- and di-*ortho* PCBs, and postnatal di-*ortho* PCB exposure. Decreased risks of infections were also indicated in infants with higher postnatal mono-*ortho* PCB and *p,p'*-DDE exposure, but the dose-response was not as clear as for postnatal di-*ortho* PCB exposure. The strong correlations between dioxin-like mono-*ortho* PCB, non-dioxin-like di-*ortho* PCB, and *p,p'*-DDE exposures among the Uppsala infants make it difficult to suggest which of the organochlorine compound groups that contributes most to the observed decline in infection risk at higher exposures. Another complicating factor is that body burdens of PCDD/Fs are strongly correlated with body burdens of mono- and di-*ortho* PCBs among the Uppsala mothers [20], making it even more difficult to draw conclusions about the contribution of non-dioxin-like and dioxin-like compounds to the observed effects.

We did not study mechanisms behind the PCB and *p,p'*-DDE modulation of infection risks, and the study was not designed to study immunoactivation or -suppression. It may however be speculated that low-dose dioxin-like effects could be involved in the seemingly protective effect of the strongly correlated mono- and di-*ortho* PCBs, and *p,p'*-DDE. Studies of TCDD-exposed rodents have indicated that parts of the immune system may be activated at low exposure levels and suppressed at higher exposures [37-39]. Moreover, pre- and postnatal exposure of rats to low doses of TCDD enhanced innate immune response to influenza virus challenge, as indicated by

neutrophilia and increased interferon γ levels in the lungs in both males and females, and suppressed cell mediated and antibody responses in females [40,41].

High prenatal CB 28+52+101 exposure increased the infection risk among the infants. Correlations between infant exposures to CB 28+52+101 and the other two PCB compound groups studied by us were weak, probably due to different sources of exposure. Food is the major source of mono- and di-*ortho* PCB exposure [42]. Our results show that food exposure to CB 28+52+101 generally is low, as indicated by the low serum lipid levels of the compounds. The high body burdens of CB 28+52+101 among some of the mothers, may be due to exposure in the in-door environment from PCB-laden building materials [28]. The associations between exposure to CB 28+52+101 could reflect other exposures common for “PCB-houses”. In our cohort, however, no associations were found between elevated concentrations of these three PCB congeners in maternal serum and residence during pregnancy in buildings built during the PCB usage period (1956-1972) [19]. It can nevertheless not be excluded that exposure to unknown environmental factors in PCB buildings contributed to the observed associations between PCB 28+52+101 exposure and risk of respiratory infections among the infants. No previous study has reported associations between infection risks among infants and pre- and postnatal exposure to CB-28, CB-52 and CB-101. No difference in health complaints was observed between children from a CB 28+52+101-contaminated school and children from a non-contaminated school [29].

The congeners CB-28, CB-52, CB-101, as well as the di-*ortho* PCBs, are non-dioxin-like congeners. Non-dioxin-like PCBs may act through pathways independent of the aryl hydrocarbon receptor, which binds dioxin-like compounds [43-46]. Little is known about immunotoxic effects of non-dioxin-like PCB congeners CB-28, CB-52 and CB-101. In contrast to the dioxin-like CB-77, CB-126 and CB -69, CB-52 induced apoptosis in mice spleen cells *in vitro* [46]. Moreover,

CB-28 and CB-52 caused rapid cell death among rat thymocytes *in vitro*, whereas CB-77 did not cause this effect at the same exposure concentration [47].

Only a few earlier studies have looked at associations between background PCB and *p,p'*-DDE exposure early in life and infant risk of respiratory infections. Among 3-6 months old Dutch and Inuit infants no significant associations were found between risks of respiratory infections and pre- or postnatal exposure to the congeners CB-118, CB-138, CB-153 and CB-180 [12, 13,48]. In one of the Inuit studies a significant positive trend was found between prenatal PCB exposure and all infections, including respiratory and gastrointestinal tract infections and otitis media [48]. No significant trends of decreased or increase risk of respiratory infections with increased *p,p'*-DDE exposure were found among the Inuit infants [13,48].

It is difficult to determine the reasons behind the diverging results between studies. Pre-and postnatal exposure levels were higher among the Dutch and Inuit infants than among infants from Uppsala. For instance, the mean level of the sum of di-*ortho* congeners CB-138, CB-153 and CB-180 was 375-620 ng/g lipid in plasma/milk from Dutch and Inuit mothers (assuming a lipid content of 0.6% of blood plasma) [12,13], whereas it was 143 ng/g lipid in blood serum among the Uppsala women. Mean level of the PCB marker congener CB-153 in cord blood/mother's plasma lipids in the other Inuit study was 2-fold higher than the mean level found in mother's serum lipids in the Uppsala cohort [19,48]. Mean body burdens of *p,p'*-DDE among Inuit mothers were 3- to 11-fold higher than among the Uppsala mothers [13,48]. Moreover, the results in the Inuit studies may have been influenced by the general health status of the infants, since acute otitis media was common among the Inuit infants [13].

Our study is small and the results should therefore be interpreted with caution. It may have been difficult for mothers to correctly report infant health history of infants. Bias could have been introduced in the diagnosis of disease, since the disease diagnosis was not confirmed from

medical records. The women did not get information about their body burdens of organochlorine compounds, avoiding bias in the reporting of diseases due to knowledge about the degree of exposure of the infant. Even though the results were adjusted with several potential confounders, unknown factors may still be involved in the associations observed. The results therefore have to be considered as hypothesis generating.

WBC and lymphocyte subset analyses were performed at the end of the 3 month study period. Therefore some of the observed associations with organochlorine compound exposure may be a consequence of the respiratory infections the infants had experienced during the study period. An increased number of lymphocytes and monocytes, as observed among infants with the highest CB 28+52+101 prenatal exposure, are indicators of infections and inflammation [49,50]. Therefore increased WBC count could be related to more recent infections. However, adjustment of the immune cell results for respiratory infections during the study period (yes/no) did not alter the observed associations. Moreover, exclusion of infants with ongoing infection at time of sampling (N=6) did not alter the results. The immune system is very complex and we only studied a few immune markers. It is consequently difficult to draw conclusions about the relation between immune cell and infection results. The few observed shifts in immune cell numbers/percentages associated with organochlorine compound exposure were generally within the normal range [51,52], making it difficult to determine the clinical consequences of the observed shifts.

Similarly as in the case of respiratory infections, prenatal exposure to mono- and di-*ortho* PCBs appeared to influence the immune cell counts in a different manner than CB 28+52+52. Mono- and di-*ortho* PCBs were negatively associated with the percentage of CD8⁺ cytotoxic T-cells. Both animal and human studies suggest that early life exposure to dioxin-like compounds may modulate the numbers/percentages of CD8⁺ cytotoxic T-cells. Among 18 months old Dutch

infants a positive association was found between CD8⁺ cell numbers and prenatal exposure to PCBs and PCDD/Fs, but no association was found at 3 months of age [12]. In mice and rat offspring the population of CD8⁺ cytotoxic T-cells was increased after pre- and postnatal TCDD exposure [2,53,54]. Differences in exposure levels of dioxin-like PCBs and PCDD/Fs may have contributed to diverging results among Uppsala (mean:19 pg total TEQ/g mother's milk lipid) and Dutch infants (64 pg TEQ/g lipid) [12,20].

We found no indications of influence of pre- and postnatal *p,p'*-DDE exposure on numbers of different types of lymphocytes, which is in accordance with the results reported for Inuit infants [13]. A negative association between prenatal *p,p'*-DDE exposure and percentage of eosinophilic granulocytes was however observed among the Uppsala infants. A study on German children in the ages 7-10 years reported a reduced eosinophilic granula content of eosinophilic granulocytes among children with the highest body burdens of *p,p'*-DDE [55]. Taken together the results indicate that eosinophilic granulocytes may respond to background exposures to *p,p'*-DDE among infants and children.

The WBC count and the analysis of lymphocyte subsets were performed on only 50-80 infants, and there were proportionally more girls in this subgroup than in the group of mother/infant pairs that did not donate blood. Moreover, fewer of the infants in the WBC group had been vaccinated. The immune cell results can therefore not be directly extrapolated to the whole study group. The immune cell numbers and percentages were only measured at one time point at the end of the 3 month study period, and we do not know if the results are representative for the other parts of the study period. Many statistical comparisons were made and it can therefore not be excluded that the results were due to chance. We used a strict significance level ($p \leq 0.01$) and the results did not change significantly after adjustment for potential confounders, which reduces the possibility of chance findings.

Conclusions

This hypothesis generating study suggest that background exposure to PCBs and *p,p'*-DDE early in life modulate immune system development. Strong correlations between mono- and di-*ortho* PCBs, and *p,p'*-DDE exposures make it difficult to identify the most important contributor to the suggested immunomodulation, and to separate effects due to pre- and postnatal exposure. Our findings may have consequences for the health development during childhood, since respiratory infections early in life may be risk factors for asthma and middle ear infections.

List of abbreviations

CB, chlorinated biphenyl; CD, clusters of differentiation; CD3⁺, T-lymphocytes; CD4⁺, T-helper cells; CD8⁺, cytotoxic cells; CD19⁺, B-lymphocytes; CD56⁺, natural killer cells; CI, confidence interval; DDE, dichlorodiphenylchloroethane; IUPAC, International Union of Pure and Applied Chemistry; PCB, polychlorinated biphenyl; SD, standard deviation; SE, standard error; WBC, white blood cell.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

AG participated in the planning of the study, data collection and data analyses, and wrote the first draft of the manuscript. AT participated in the planning of the study, data collection and data analyses. MA participated in the planning of the study and was responsible for the organochlorine compound analysis. AJ participated in the planning of the study and data analyses, and was responsible for the lymphocyte subset analyses. POD participated in the planning of the study and data collection. GR was responsible for the WBC analyses. SC participated in the planning of the study and data collection. All authors participated in the preparation of the final manuscript and approved the submission.

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Figure legends

Figure 1. White blood cells (WBC) and prenatal exposure to PCB congeners CB-28, CB-52 and CB-101. Unadjusted and adjusted means (\pm SE) of WBC numbers and percentages in three months old infants prenatally exposed to the sum of CB-28, CB-52 and CB-101. Lipid adjusted serum concentrations of the PCB compounds measured in the blood of the mothers in late pregnancy (week 32-34) were used as a measure of prenatal exposure. Adjusted means were calculated in cases when statistically significant results were found in the unadjusted analysis. Results were adjusted for age of the mother, smoking and alcohol during pregnancy, mother's education, vaccination of the infant, nursing of the infant, age of the infant, infant's respiratory infections. The exposure variable was categorized since over 40% of the study participants had serum lipid levels of the PCBs below the limit of quantification. An effort was made to have equal numbers of participants in the two other exposure categories. *Significantly different from the group with the lowest exposure (reference category) (N= 81-85, $p \leq 0.01$).

Figure 2. Lymphocyte subsets and prenatal exposure to the PCB congeners CB-28, CB-52 and CB-101. Unadjusted means (\pm SE) of numbers and percentages of lymphocyte subsets in three months old infants prenatally exposed to the sum of CB-28, CB-52 and CB-101. Lipid adjusted serum concentrations of the PCB compounds, measured in the blood of the mothers in late pregnancy (week 32-34), were used as a measure of prenatal exposure. $CD4^+/CD8^+$: the ratio between numbers of $CD4^+CD8^-$ and $CD4^-CD8^+$ lymphocytes. The exposure variable was categorized since over 40% of the study participants had serum lipid levels of the PCBs below the limit of quantification (exposure category 1). An effort was made to have equal numbers of

participants in the two other exposure categories. *Significantly different from the group with the lowest exposure (reference category) (N=47-52, $p \leq 0.01$).

Table 1. Characteristics of the participating mother/child pairs^a.

Variable	Study groups			White blood cells (N=86)			Lymphocyte subsets (N=52)		
	Infections (N=190)			White blood cells (N=86)			Lymphocyte subsets (N=52)		
Mother's age (yr)	28 (21-41)			29 (21-36)			29 (22-35)		
Infant's age (d)	92 (75-123)			93 (76-112)			93 (76-112)		
	Percent			Percent			Percent		
School yrs	≤13:50	14-16:24	>16:26	≤13:50	14-16:27	>16:23	≤13:43	14-16:30	>16:27
Smoking ^b	No:82	Yes:18		No:86	Yes:14		No:89	Yes:11	
Alcohol ^b	No:82	Yes:18		No:83	Yes:17		No:77	Yes:23	
Nursing ^c	Whole:81	Partial:19		Whole:84	Partial:16		Whole:91	Partial:9	
Infant's sex	Girl:44	Boy:56		Girl:58	Boy:42		Girl:57	Boy:43	
Vaccination	No:69	Yes:31		No:76	Yes:24		No:85	Yes:15	
Resp. infection	No:72	Yes:28		No:71	Yes:29		No:78	Yes:22	

^aMother's and infant's age: median (range). WBC were analyzed in a subgroup of 86 infants, and within this subgroup lymphocyte subsets were analyzed among 52 infants with enough blood left after WBC count analysis.

^bDuring pregnancy.

^cPartial nursing also includes no nursing.

Table 2. Organochlorine concentrations in mother's serum lipids in late pregnancy and infant exposure from mother's milk^a.

Exposure	Study groups		
Serum ^b	Infections ^d	White blood cells ^d	Lymphocyte subsets ^d
CB 28+52+101	4 (3-427)	4 (3-427)	4 (3-19)
Di-ortho PCB	131 (44-362)	127 (44-342)	129 (44-342)
Mono-ortho PCB TEQ	4 (1-20)	4 (1-11)	4 (1-9)
<i>p,p'</i> -DDE	88 (21-622)	85 (24-622)	83 (29-622)
Mother's milk exposure ^c			
CB 28+52+101	8 (0-155)	8 (0-76)	9 (0-39)
Di-ortho PCB	364 (0-1666)	351 (0-830)	372 (0-830)
Mono-ortho PCB TEQ	11 (0-54)	11 (0-26)	11 (0-26)
<i>p,p'</i> -DDE	311 (0-2199)	289 (0-2199)	306 (0-2199)

^aMedian (range). Di-ortho PCB=CB-138, CB-153, CB-180; Mono-ortho PCB TEQ=CB-105, CB-118, CB-156, CB-167 [56]. The study group "Infections" consists of all study participants, the "White blood cells" group is a subgroup of infants among the study participants which had their WBC counts analyzed, and within this subgroup lymphocyte subsets were analyzed in the "Lymphocyte" group composed of infants with enough blood left after WBC count analysis.

^bng/g lipid. Mono-ortho PCBs: pg TEQ/g lipid. Serum sampled week 32-34.

^cng/g fresh weight*days. Mono-ortho PCBs: pg TEQ/g fresh weight*days. Calculated as organochlorine concentration in mother's milk on a fresh weight basis*days of nursing*(%of full nursing/100).

^dInfections:Prenatal exposure, N=190; Postnatal exposure, N=175. Differential count:N=86. Lymphocyte subsets:N=52.

Table 3. Numbers and percentages of white blood cells (WBC) and lymphocyte subsets in 3 month-old infants^a.

Differential count	N	No. of cells x 10 ⁹ /L	N	% of WBC
White blood cells	81	8 (5-15)		
Neutrophils	80	1.6 (0.6-5.7)	85	21 (8-47)
Eosinophils	80	0.3 (0.1-1.0)	85	4 (0.5-10)
Lymphocytes	80	5.4 (2.9-9.5)	85	70 (37-86)
Monocytes	80	0.3 (0.1-1.4)	85	4 (1-15)
Lymphocyte subsets				% of lymphocytes
CD19 ⁺	47	0.8 (0.1-2.0)	51	16 (2-34)
CD3 ⁺	47	3.7 (1.5-7.3)	52	70 (38-85)
CD4 ⁺ CD8 ⁻	47	3.1 (1.0-5.3)	52	55 (24-72)
CD4 ⁻ CD8 ⁺	47	0.7 (0.2-2.1)	52	13 (5-23)
CD56 ⁺	47	0.05 (0-0.1)	52	0.8 (0-2.2)

^aMedian (range)

Table 4. Regression coefficients for associations between organochlorine exposure prenatally and numbers and percentages of numbers white blood cells and lymphocyte subsets^a

	Di-ortho PCB	Mono-ortho PCB TEQ	<i>p,p'</i> -DDE
White blood cell count ^b	-0.01±0.54	0.28±0.44	0.49±0.36
Neutrophil numbers ^b	0.04±0.18	0.08±0.14	-0.03±0.12
Neutrophil % ^b	0.99±2.10	0.33±1.73	-0.74±1.47
Eosinophil numbers ^b	0.01±0.04	0.02±0.03	-0.06±0.03
Eosinophil % ^b	0.09±0.55	0.06±0.45	-1.04±0.36*
Eosinophil % ^c			-1.68±0.43*
Lymphocyte numbers ^b	-0.06±0.42	0.14±0.35	0.50±0.28
Lymphocyte % ^b	-0.93±2.23	0.08±1.84	1.33±1.56
Monocyte numbers ^b	0.02±0.05	0.001±0.04	0.04±0.03
Monocyte % ^b	0.59±0.50	0.16±0.42	0.41±0.35
CD19 ⁺ numbers ^b	-0.04±0.16	-0.05±0.15	0.02±0.11
CD19 ⁺ % ^b	0.05±2.6	0.87±2.3	-0.81±1.7
CD3 ⁺ numbers ^b	-0.02±0.46	-0.14±0.42	0.42±0.30
CD3 ⁺ % ^b	-3.4±4.0	-3.6±3.5	-1.2±2.8
CD4 ⁺ CD8 ⁻ numbers ^b	0.14±0.36	0.06±0.33	0.29±0.24
CD4 ⁺ CD8 ⁻ % ^b	0.01±3.78	-0.16±3.37	-0.61±2.61
CD4 ⁻ CD8 ⁺ numbers ^b	-0.24±0.09	-0.19±0.08	-0.15±0.06
CD4 ⁻ CD8 ⁺ % ^b	-4.02±1.29*	-3.35±1.14*	-2.25±0.98
CD4 ⁻ CD8 ⁺ % ^c	-5.61±1.96*	-6.24±1.84*	
CD4 ⁺ /CD8 ⁺ ^b	0.91±0.61	0.71±0.55	0.44±0.41
CD56 ⁺ numbers ^b	0.01±0.01	0.003±0.008	0.003±0.003
CD56 ⁺ % ^b	0.19±0.18	0.11±0.16	0.06±0.12

^aPrenatal exposure: Lipid adjusted mother's serum organochlorine compound concentrations in late pregnancy (week 32-34). Di-ortho PCB=CB-138, CB-153, CB-180; Mono-ortho PCB TEQ=CB-105, CB-118, CB-156, CB-167 [56]. White blood cell counts: N=80-85. Lymphocyte subsets: N=47-52.

^bRegression coefficients from simple regression analysis (mean±SE)

^cPartial regression coefficients (mean±SE) adjusted for age of the mother, smoking and alcohol during pregnancy, mother's education, vaccination of the infant, nursing of the infant, age of the infant, and infant's respiratory infections.

***p≤0.01.**

Table 5. Odds ratios (95% CI) for associations between respiratory infection during the first 3 months after birth and pre- or postnatal exposure to organochlorine compounds^a

	Model ^b	Exposure categories ^c			
		Category 1	Category 2	Category 3	Category 4
Prenatal					
CB 28+52+101	Unadjusted	1.0	1.5 (0.66-3.3)	2.6 (1.2-5.6)	
	Multivariate	1.0	1.7 (0.71-4.1)	3.4 (1.4-7.8)	
Di-ortho PCB	Unadjusted	1.0	0.35 (0.14-0.86)	0.34 (0.14-0.83)	0.49 (0.21-1.2)
	Multivariate	1.0	0.28 (0.10-0.79)	0.23 (0.07-0.71)	0.29 (0.08-1.0)
Mono-ortho PCB TEQ	Unadjusted	1.0	0.71 (0.30-1.7)	0.54 (0.23-1.3)	0.46 (0.19-1.2)
	Multivariate	1.0	0.58 (0.22-1.6)	0.34 (0.10-1.1)	0.23 (0.06-0.91)
<i>p,p'</i> -DDE	Unadjusted	1.0	0.67 (0.27-1.6)	0.73 (0.30-1.78)	0.80 (0.34-1.9)
	Multivariate	1.0	0.64 (0.24-1.7)	0.69 (0.25-1.9)	0.74 (0.25-2.2)
Postnatal					
CB 28+52+101	Unadjusted	1.0	0.51 (0.19-1.4)	0.72 (0.28-1.9)	1.4 (0.58-3.4)
	Multivariate	1.0	0.44 (0.14-1.4)	0.65 (0.23-1.8)	1.4 (0.53-3.8)
Di-ortho PCB	Unadjusted	1.0	0.48 (0.19-1.2)	0.53 (0.21-1.3)	0.31 (0.12-0.83)
	Multivariate	1.0	0.27 (0.08-0.87)	0.26 (0.08-0.85)	0.14 (0.04-0.50)
Mono-ortho PCB TEQ	Unadjusted	1.0	0.36 (0.14-0.97)	0.53 (0.21-1.3)	0.69 (0.29-1.7)
	Multivariate	1.0	0.23 (0.07-0.79)	0.37 (0.12-1.2)	0.33 (0.10-1.1)
<i>p,p'</i> -DDE	Unadjusted	1.0	0.25 (0.09-0.71)	0.53 (0.21-1.3)	0.69 (0.29-1.7)
	Multivariate	1.0	0.18 (0.06-0.60)	0.40 (0.14-1.2)	0.52 (0.17-1.5)

^aPrenatal exposure: Lipid adjusted mother's serum organochlorine compound concentrations in late pregnancy (week 32-34). Postnatal exposure: mother's milk concentrations on a fresh weight basis*days of nursing*(%of full nursing/100). Di-ortho PCB=CB-138, CB-153, CB-180; Mono-ortho PCB TEQ=CB-105, CB-118, CB-156, CB-167 [56]. Prenatal exposure:N=190. Postnatal exposure:N=175. **Bold odds ratios** significantly different from the odds ratio of the reference category (p≤0.05).

^bMultivariate model also included the independent variables age of the mother, smoking and alcohol during pregnancy, mother's education, vaccination of the infant, nursing of the infant, and age of the infant.

^cPre- and postnatal exposure was categorized with the lowest exposure in Category 1 and increasing exposure with increasing Category number. Efforts were made to have equal numbers of participants in the exposure categories for each compound group.

Additional files

File name: Table A, Table B, Table C, Table D, Table E

File format: MS Word document.

Title of data:

Respiratory infections and white blood cell and lymphocyte subset numbers/percentages in 3 months old infants exposed to PCB and *p,p'*-DDE postnatally.

Description of data:

5 Tables

Table A. Unadjusted means (\pm SE) of white blood cell numbers ($\times 10^9$) in 3-month-old infants exposed to organochlorines postnatally

Table B. Unadjusted means (\pm SE) of white blood cell percentages in 3-month-old infants exposed to organochlorines postnatally

Table C. Unadjusted means (\pm SE) of lymphocyte subset numbers ($\times 10^9$) in 3-month-old infants exposed to organochlorines postnatally

Table D. Unadjusted means (\pm SE) of lymphocyte subset percentages in 3-month-old infants exposed to organochlorines postnatally

Table E. Odds ratios (95% CI) for associations between respiratory infections, lasting 3 days or more during the first 3 months after birth, and pre- or postnatal exposure to organochlorine compounds

Figure 1

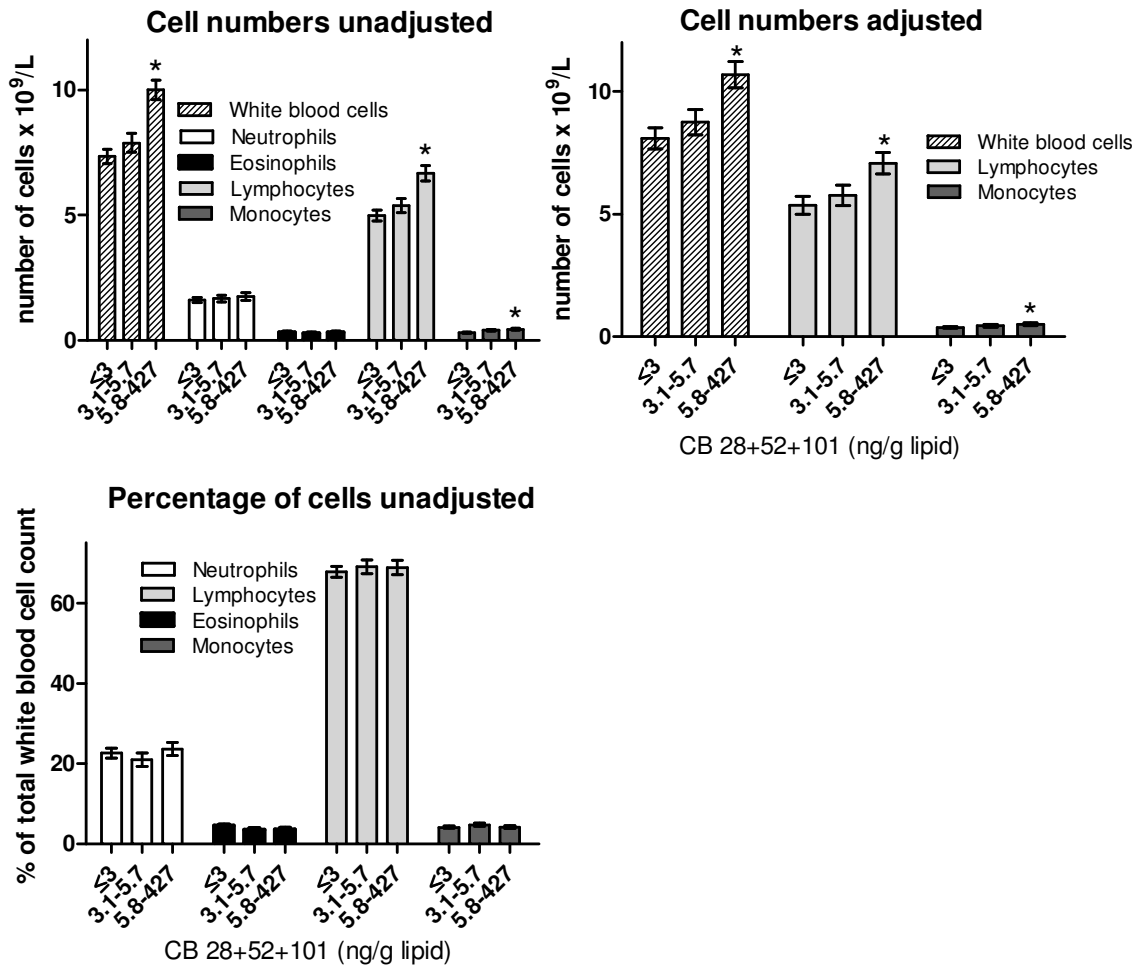
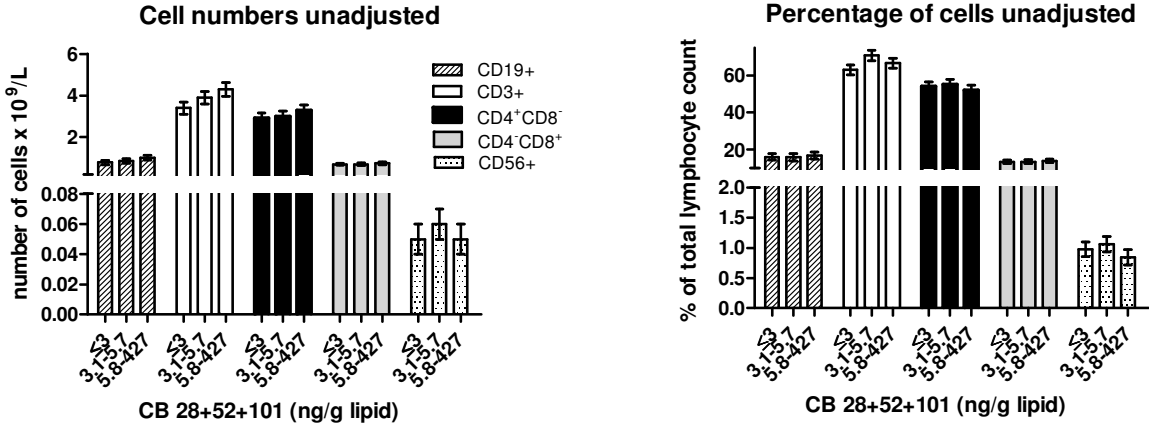


Figure 2



Additional files provided with this submission:

Additional file 1: table a.pdf, 17K

<http://www.ehjournal.net/imedia/2067971103233954/supp1.pdf>

Additional file 2: table b.pdf, 17K

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